Advanced diagnostic methods in hematology

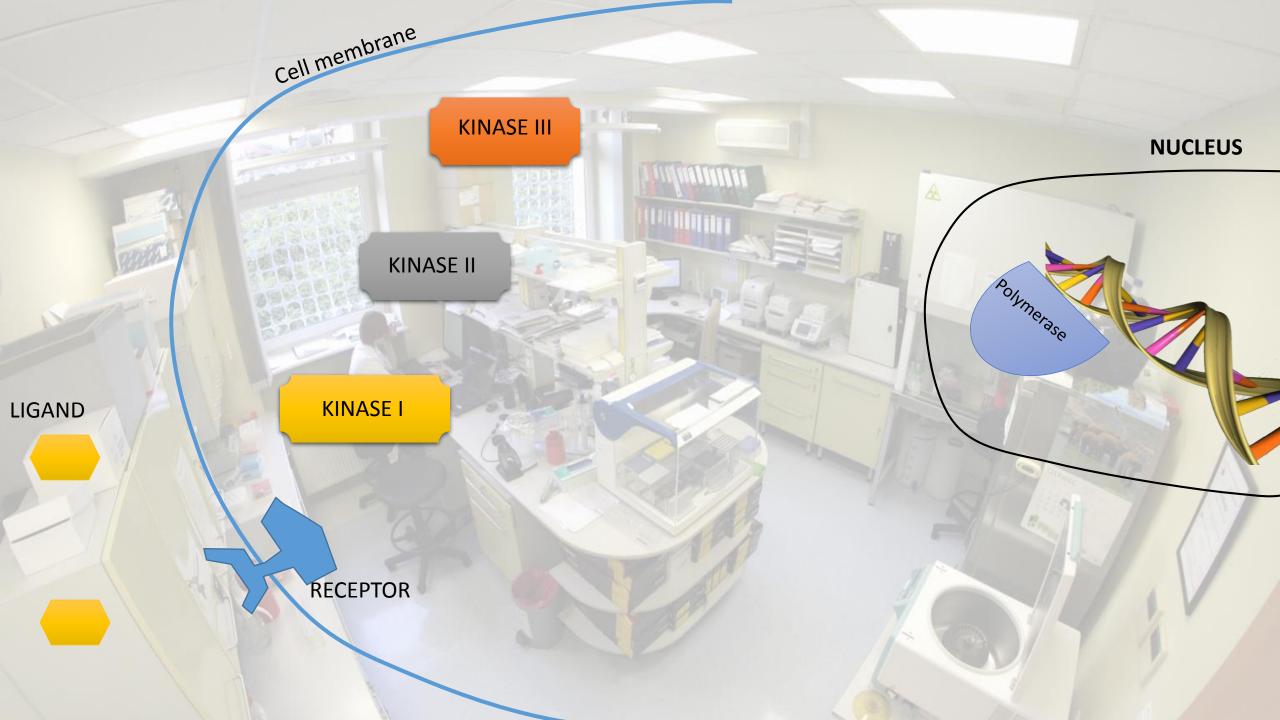
Michal Gniot Department of Hematology Poznan University of Medical Sciences

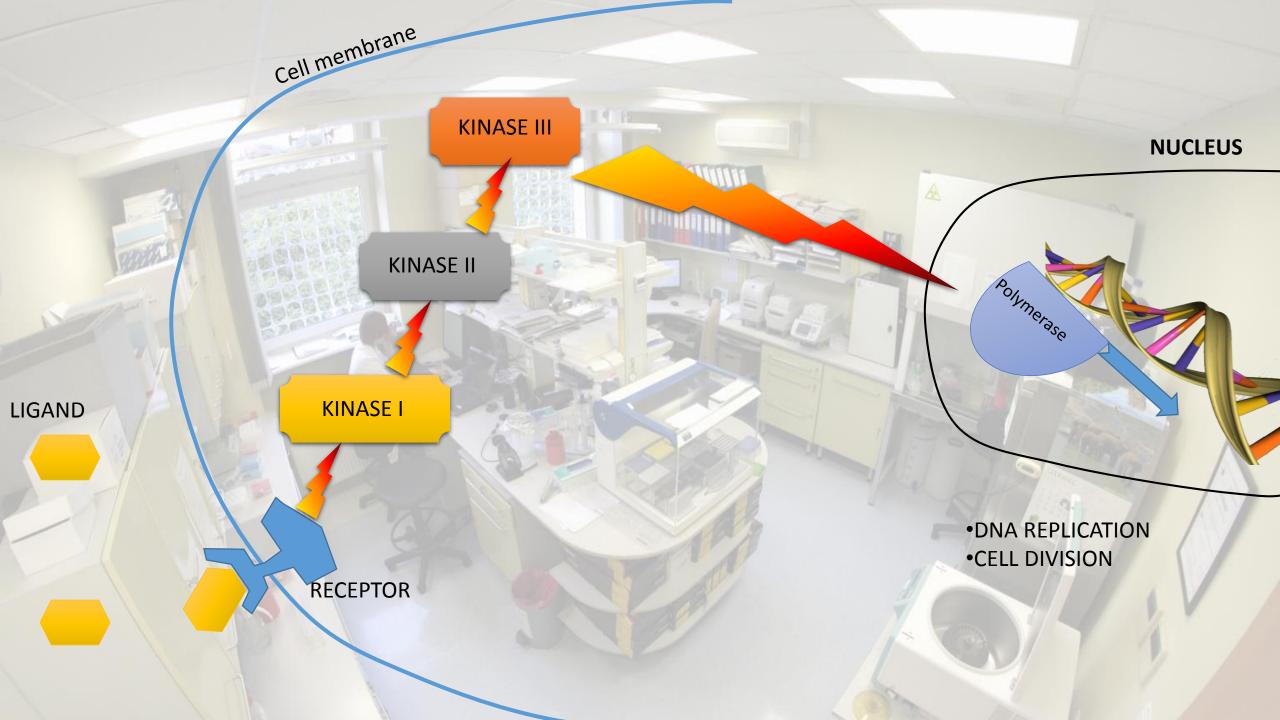
Hematological neoplasms

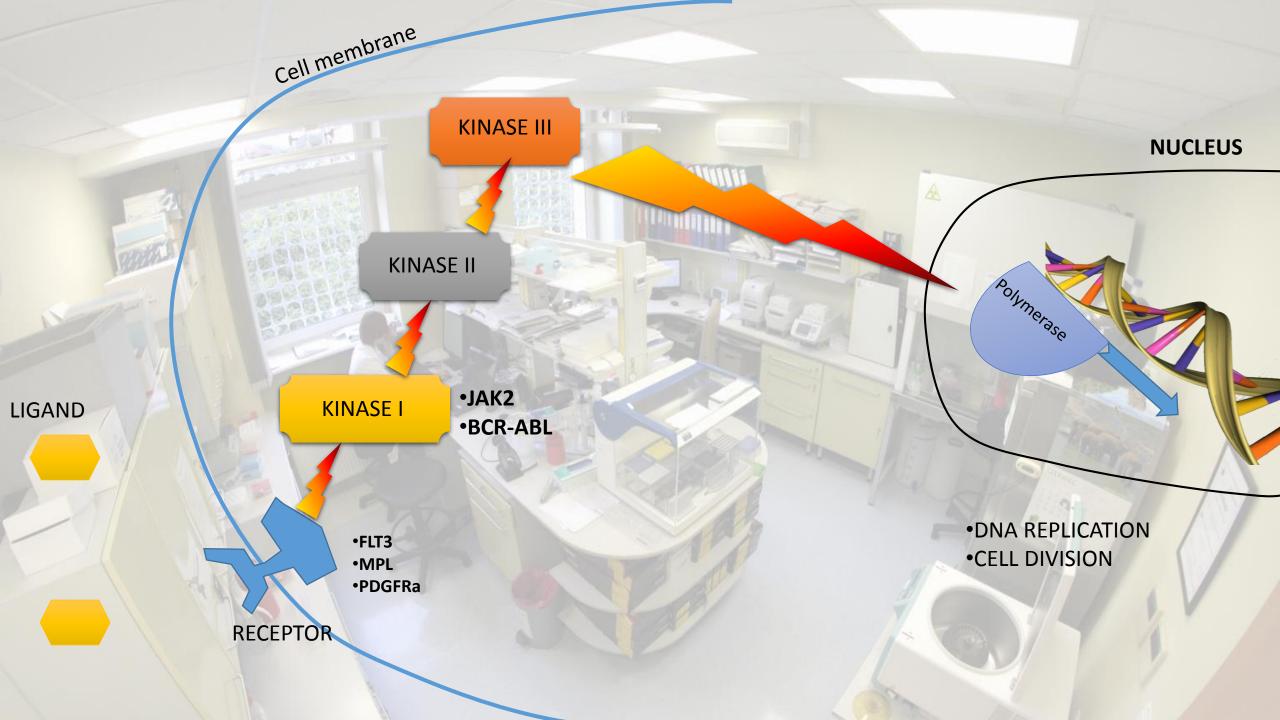
- Clonal growth of cell population originating from myeloid or lymphoid lineage
 - Somatic mutation
 - Neoplastic cells circulating in peripheral blood
 - Possible abnormalities in cell maturation
- Examples:
 - Leukemias (acute and chronic, myeloid and lymphoid)
 - Myeloproliferative neoplasms (Polycythemia Vera, Idiopatic Myelofibrosis, Essential Thrombocythemia)
 - Lymphomas, myelomas

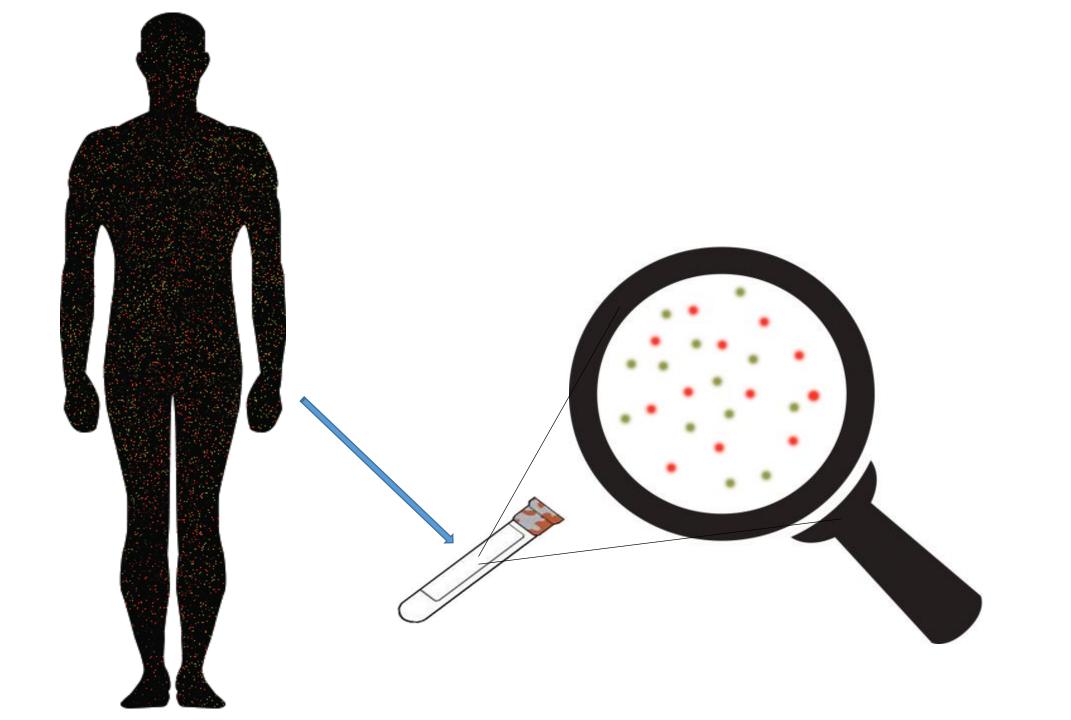
Hematological neoplasms – background

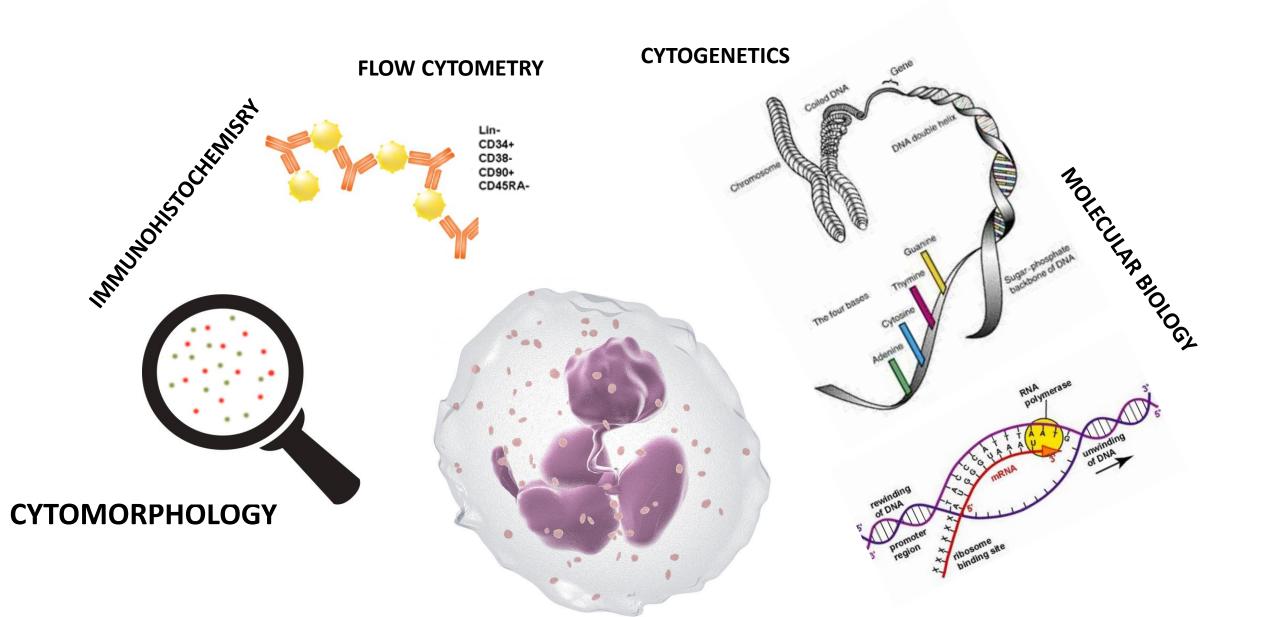
- Somatic (accquired) mutation
 - Region encoding a protein which plays a role in cell signaling pathway
 - Change of function (usually activation)
 - Point mutations, insertions-deletions, translocations
 - Translocations result in fusion genes



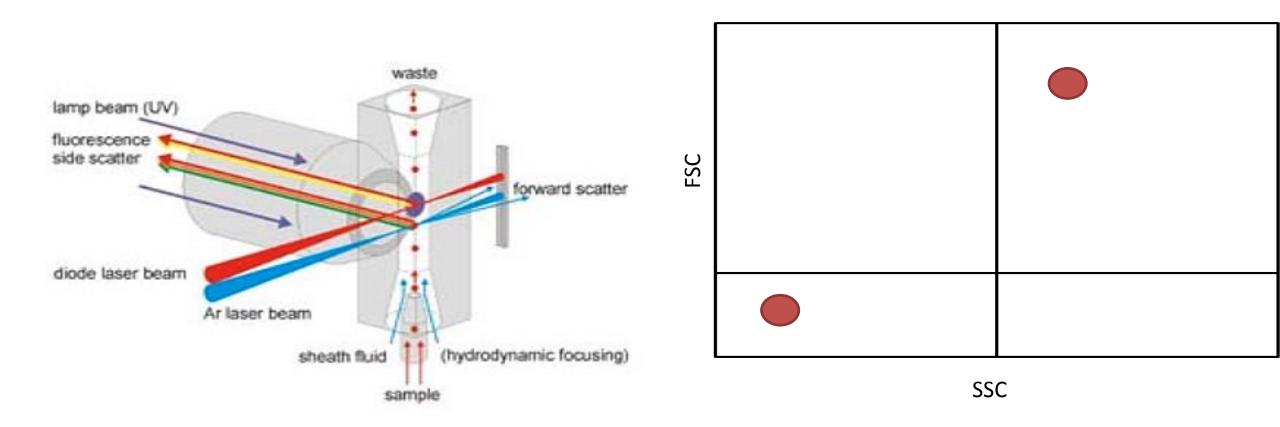




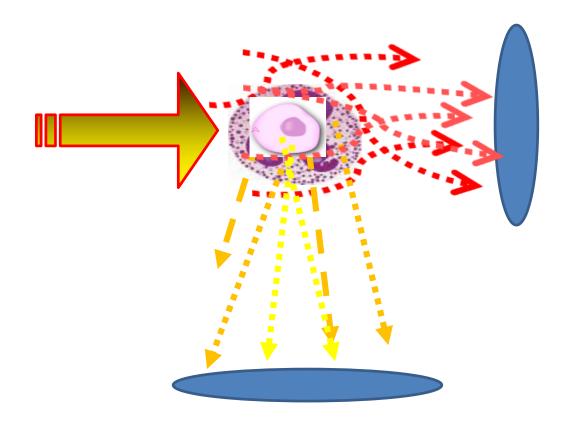




Flow cytometry

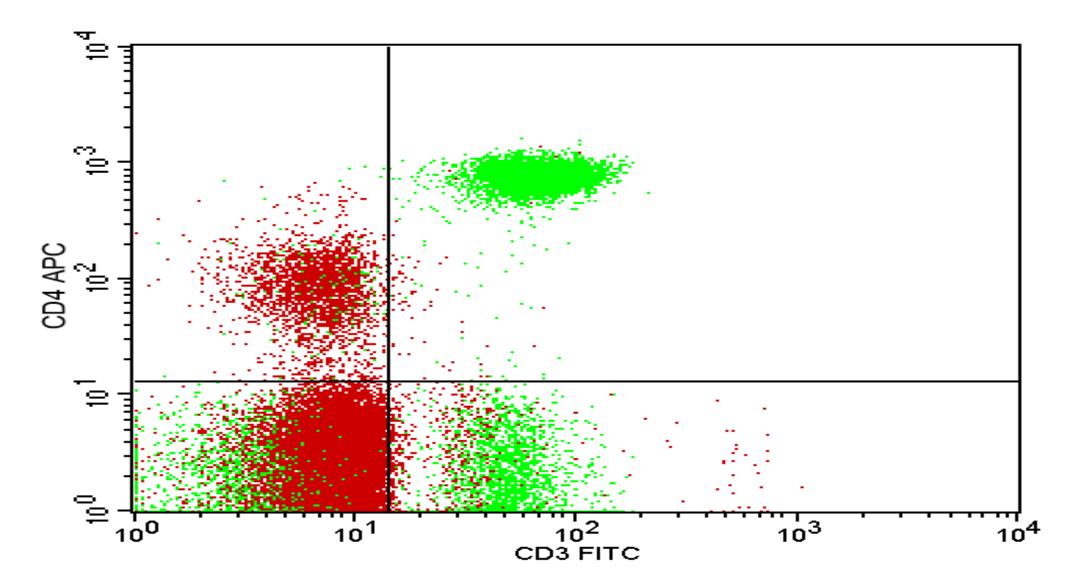


Flow cytometry

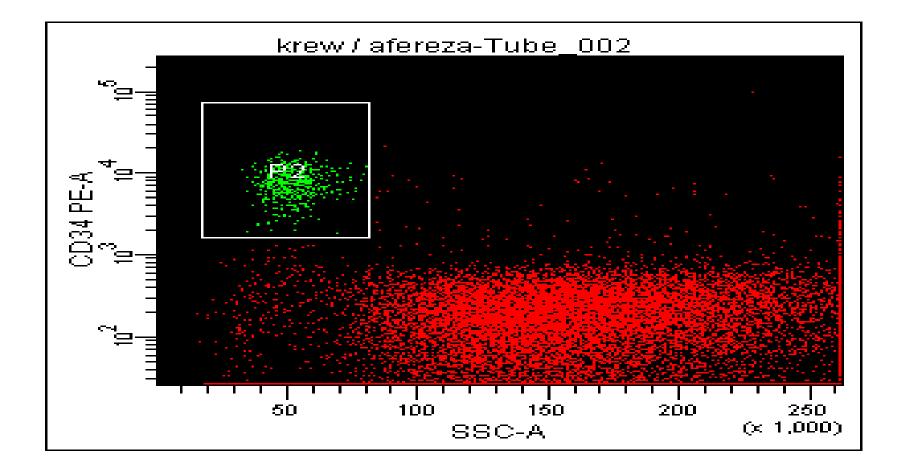


- Characterisation of hematopoietic cells in blood, marrow, CSF, exudates
- Immunophenotype describing
- Separation of cells with specific antigenity (e.g. CD34+ stem cells for transplatations)

Examples: T-helper lymphocytes (CD3+ CD4+)



CD34+ cells



	MYELOID L	INEAGE	LYMPHOID LINEAGE			
Stem cells CD34 CD117 Pre-myeloid CD33 MPO	Granulocyte CD10 CD11b CD11c CD13 CD14 CD59 CD61 CD62L CD64 CD66 CD68 MPO Megakaryocyt Platelet CD61 CD61 CD61 CD61 CD71	CD1-	CD2 CD3 CD4 CD5 CD7 CD8 CD38 CD45 CD54 CD54 CD56 CD57 CD59 CD61 CD62L CD69 CD61 CD62L CD69 CD71	B cell CD5 CD10 CD19 CD20 CD38 CD40 CD45 CD54 CD54 CD59 CD61 CD62L NK Cel CD2 CD11b CD11c CD56 CD57 CD59 CD61 CD59 CD61 CD62L		

MARKER	DESCRIPTION				
CD3	Pan T-Cell				
CD4	Helper T-Cell				
CD8	Cytotoxic/Suppressor T-Cell				
CD19	Early B-Cell, B-Cell Specific				
CD33	Early Myeloid Cells				
CD34	Stem Cells, Progenitor Cells				
CD38	Activated T-Cells, Early Progenitor Cells				
HLA-DR	Activated T-Cell, Monocytes, B-Cell				

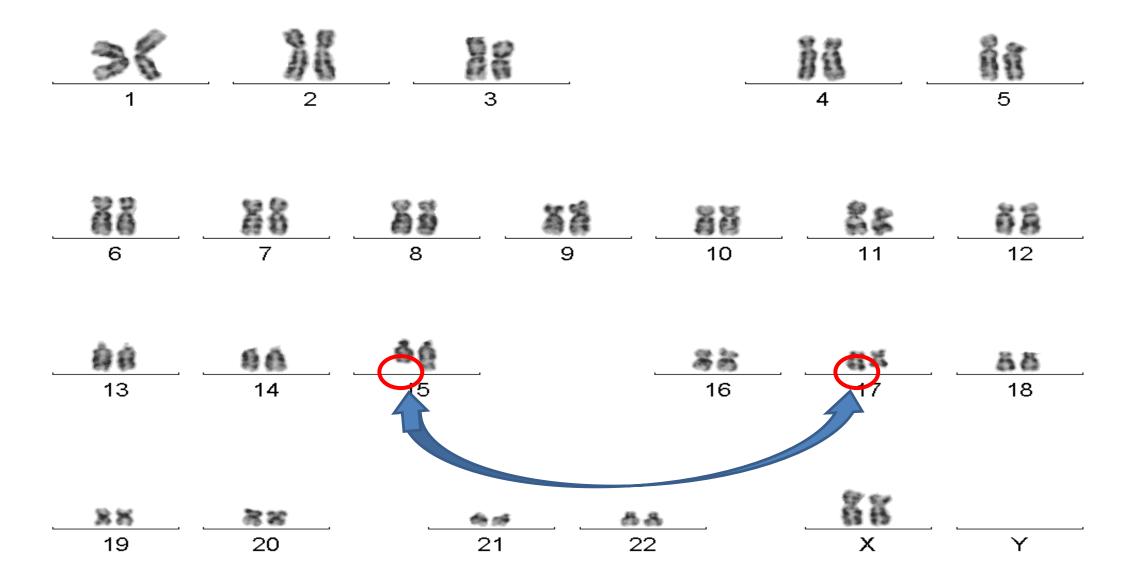
Cytogenetics

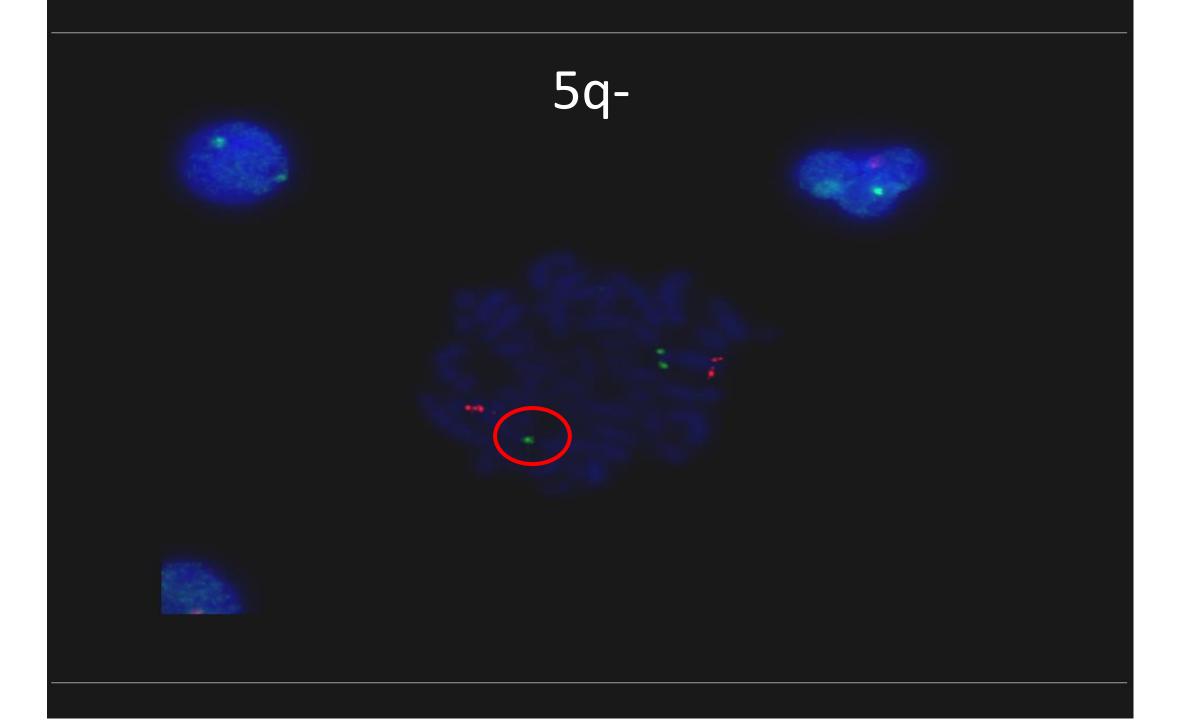
- Classical cytogenetics require culturing of bone marrow cells in order to obtain nuclei in metaphase stage.
- The chromosomes are stained using GTG (or other) banding method, photographed and analyzed in dedicated software
- Usually, 20-30 metaphases has to be assessed in comparison to normal human karyotype
- Molecular cytogenetics, FISH method (Fluorescent In-Situ Hybrydization) is a supporting method, which allows to detect specific aberrations

Aims of cytogenetic analyses

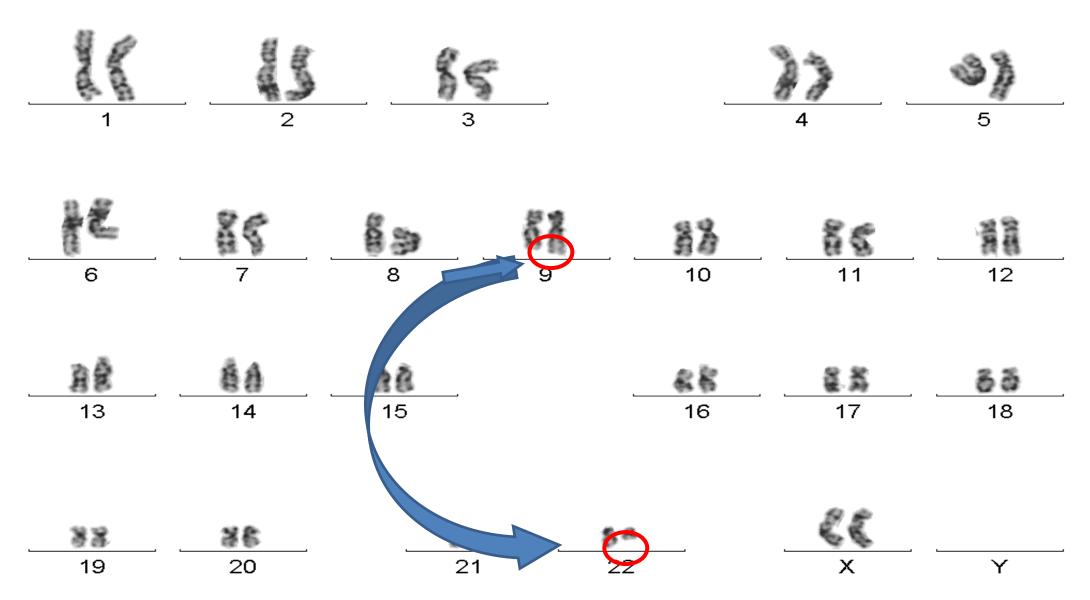
- Detection of chromosomal abnormalities characteristic to: MDS, AML, ALL, CML ...
- Observation of new, acquired abnormalities (clonal evolution)
- Prognostic factors (i.e. additional aberrations in CML)
- Monitoring of the therapy
- Post-transplantation chimerism of Y-X chromosomes

t(15;17) – PML-RARA in AML M3





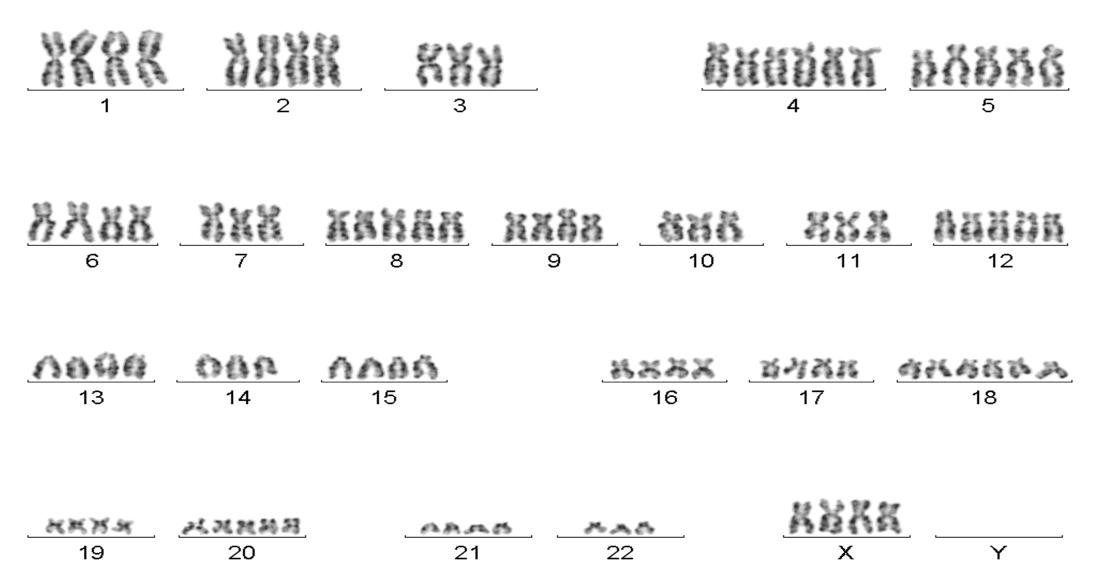
t(9;22) – Philadelphia chromosome





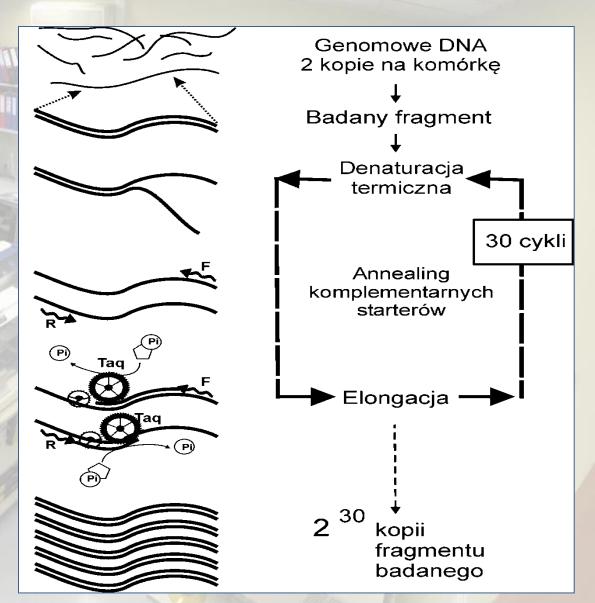


Polyploidy



Molecular biology

- Detection of specific molecular abnormalities
 - Fusion genes
 - Point mutations
 - In-del mutations
- Treatment monitoring
- Prognostic factors
- Most sensitive method for MRD monitoring*
- Post-transplantation chimerism



Main areas of interest

Diagnose

Clasification criteria

Prognostic factors

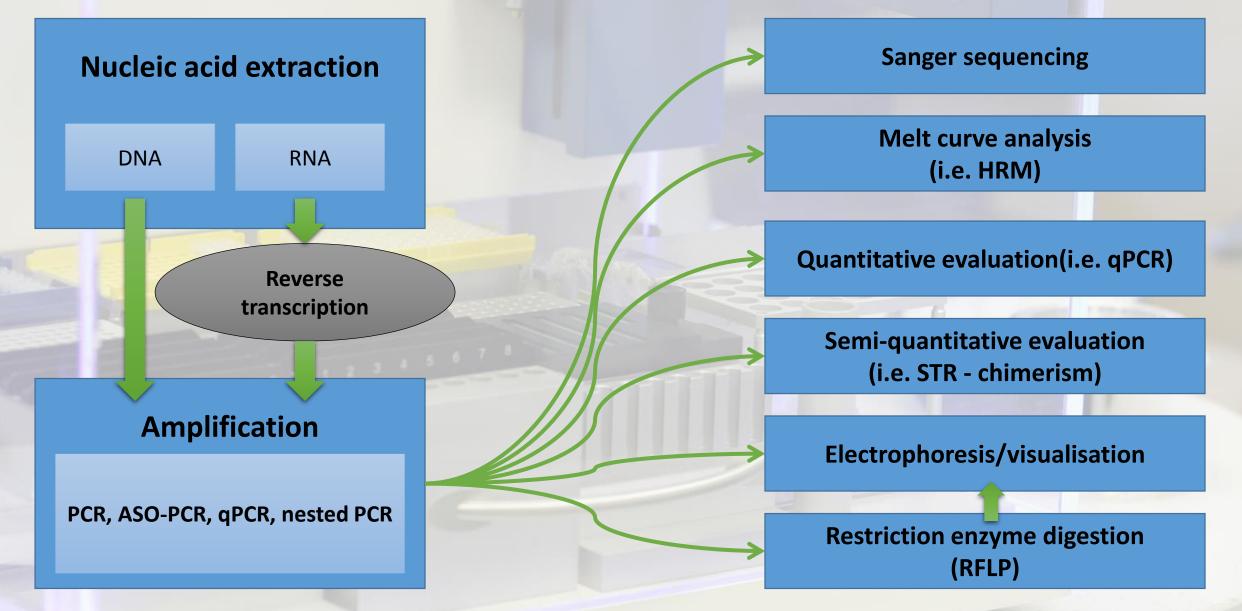
Identification of molecular targets

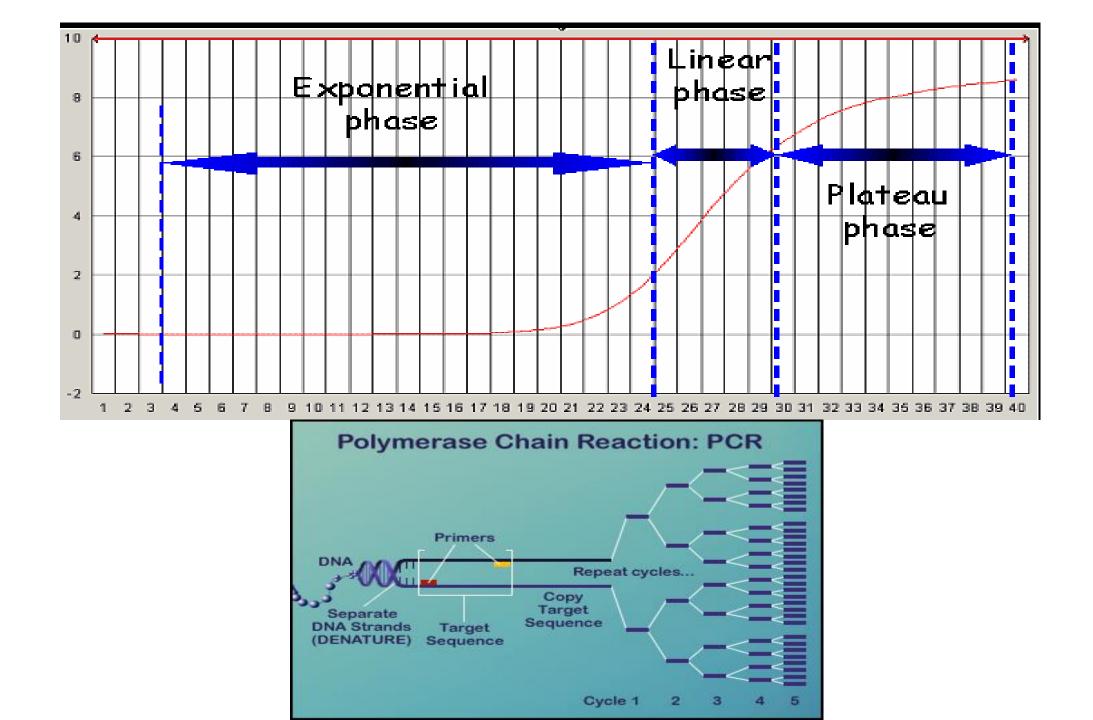
Monitoring

Based on a specific genetic marker detected at diagnosis

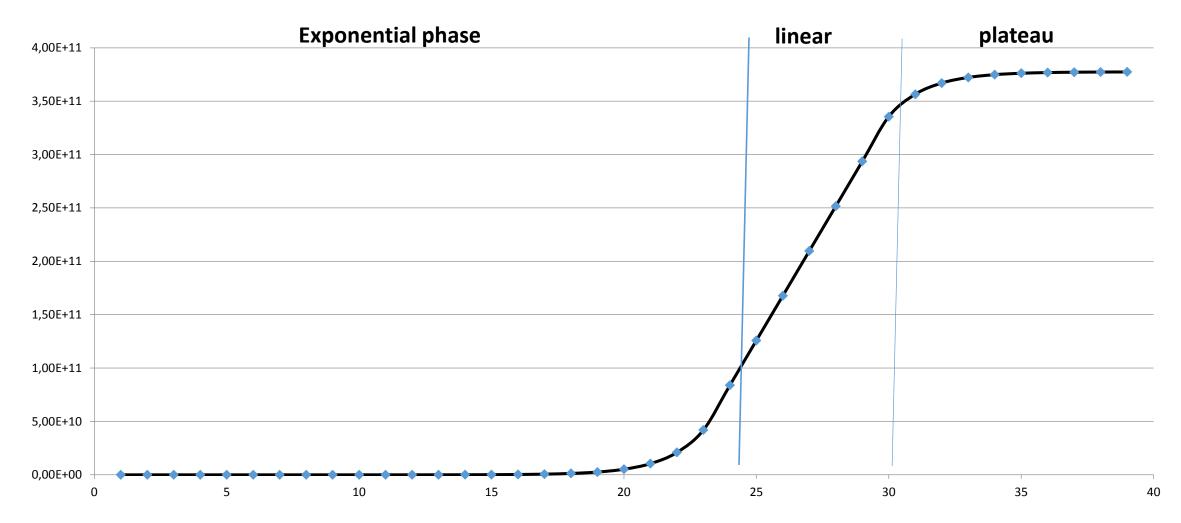
Chimerism (donor vs recipient comparison)

Molecular biology strategies

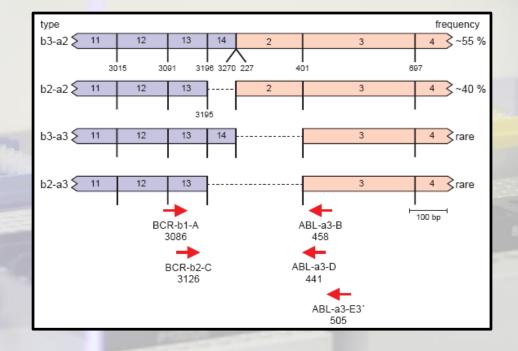


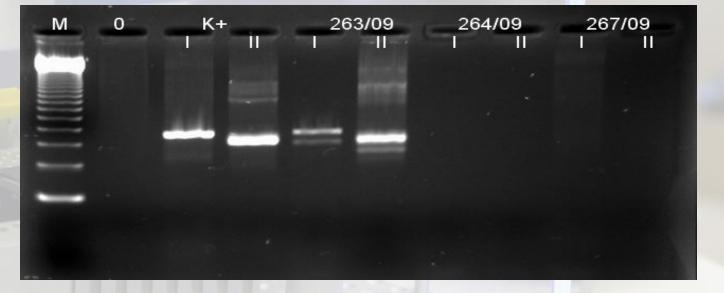


Kinetics of PCR

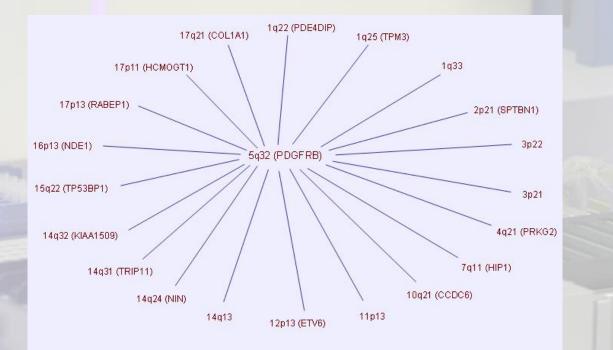


Fusion gene detection

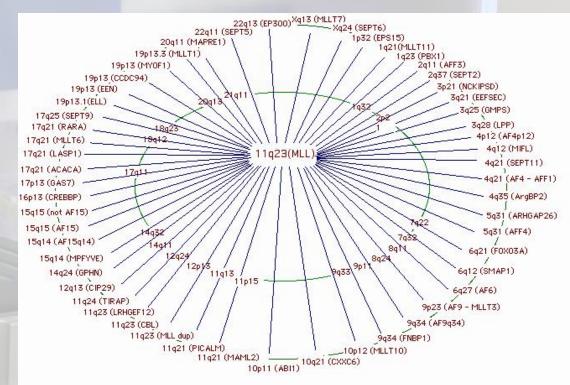




Fusion gene detection



PDGFRB and 20 partners and/or recurrent translocations. Editor 08/2004; last update 02/2009



MLL and partners - 73 recurrent translocations and 54 partner genes. Editor 06/2000; last update 10/2007

Fusion genes:

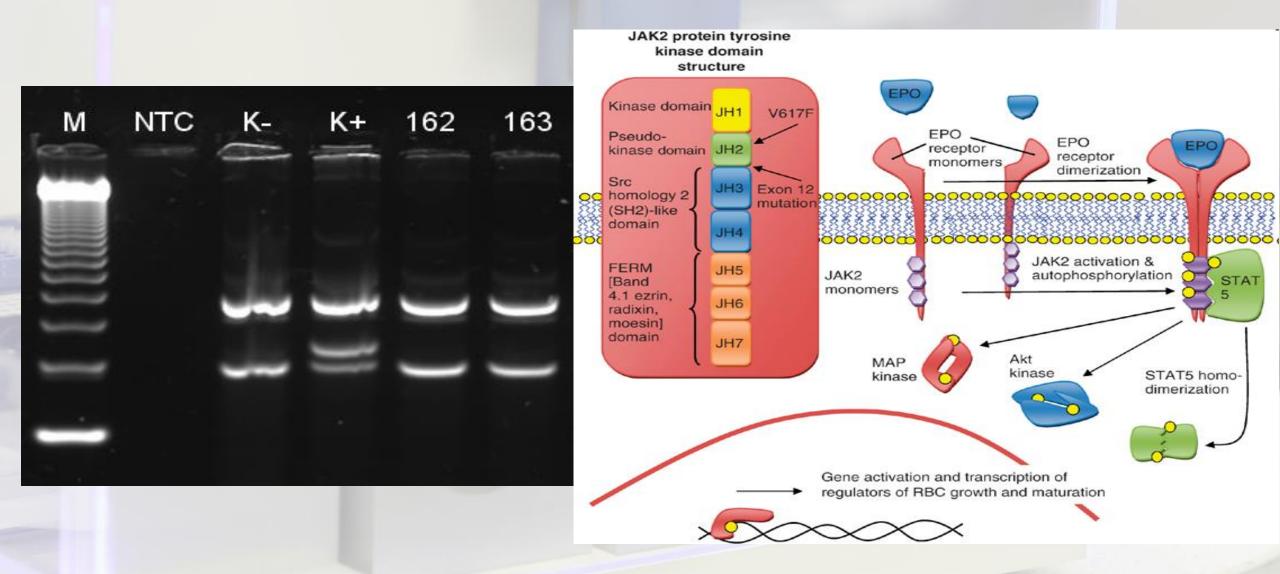
- TEL-AML1 (PCR)
- E2A-PBX (PCR)
- MLL rearangements (multiplex PCR)
- SIL-TAL (PCR)
- BCR-ABL (multiplex PCR)
- AML1-ETO (PCR)
- PML-RARA (PCR)
- CBFB-MYH11 (PCR)
- FIP1L1-PDGFRA (PCR)

Other mutations:

- JAK2 V617F (ASO QPCR)
- JAK2 ex12 (HRM/CE/SEQ)
- MPL (HRM/SEQ)
- CALR (HRM/CE/SEQ)
- FLT3 ITD (CE)
- FLT3 TKD (RFLP)
- BRAF V600E (ARMS PCR)
- CEBPA (SEQ)
- Post transplantation chimerism (CE)

To be introduced: ASXL1, LNK, CS3FR

JAK2 V617F mutation detection



JAK2 V617F standarization



Certificate of Participation

We hereby certify that:

Pracownia Biologii Molekularnej Laboratorium Diagnostyki Hematologicznej Poznań, Poland

Participated in the 2012 COST MPN&MPNr-EuroNet JAK2-V617F Inter-laboratory Quality Control Study



Niels Pallisgaard, Vejle, Denmark Julia Asp, Göteborg, Sweden Sylvie Hermouet, Nantes, France



Certificate of Participation

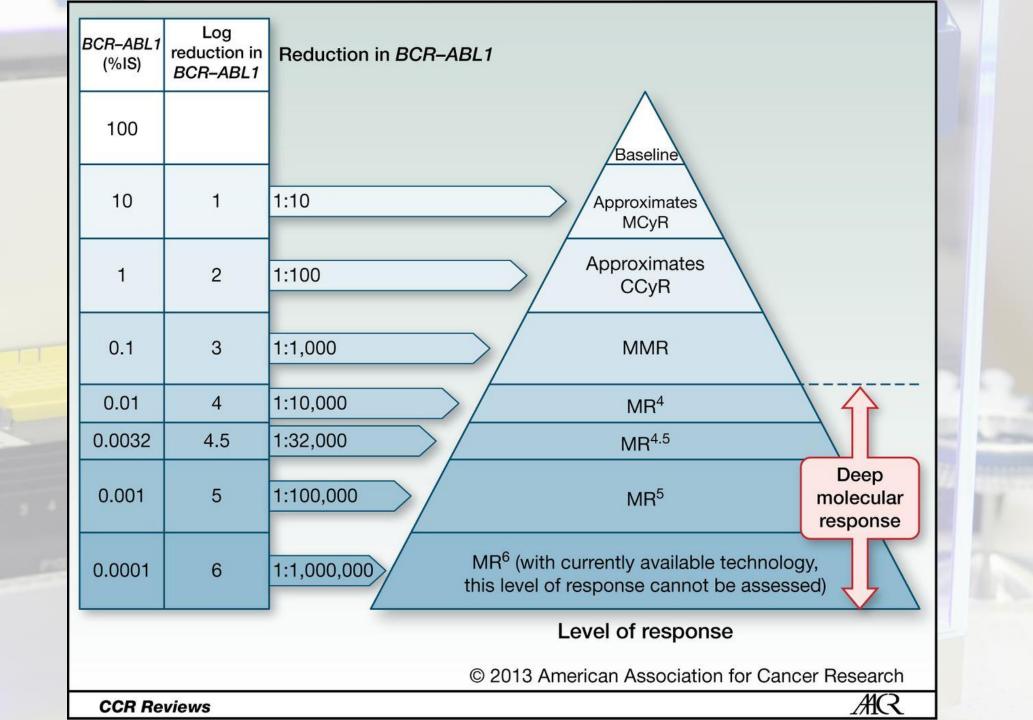
We hereby certify that:

Laboratorium Diagnostyki Hematologicznej Klinika Hematologii i Transplantacji Szpiku Poznán, Polen

participated in the 2015 MPN&MPNr-EuroNet JAK2 V617F Inter-laboratory Quality Control

> Niels Pallisgaard, Roskilde, Denmark Julia Asp, Göteborg, Sweden Sylvie Hermouet, Nantes, France

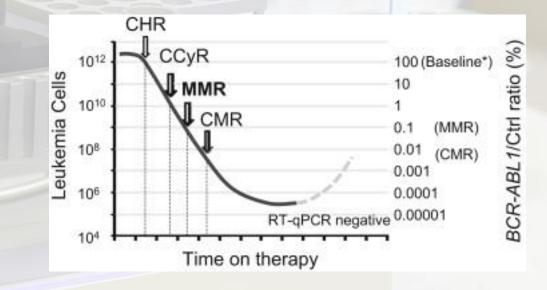




CML – recommendations based on molecular and cytogenetic analyses

OPTIMAL RESPONSE BCR-ABL trancript level below:

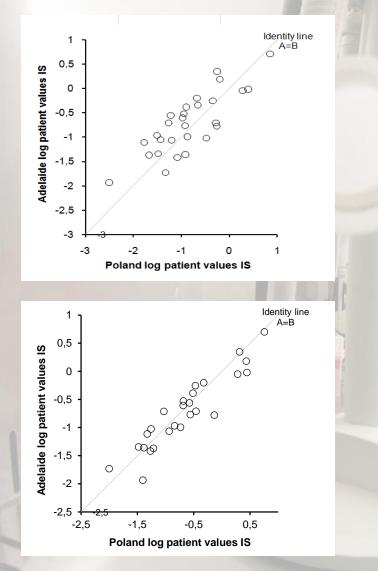
10% after 3 months1% after 6 months0,1% after 12 months

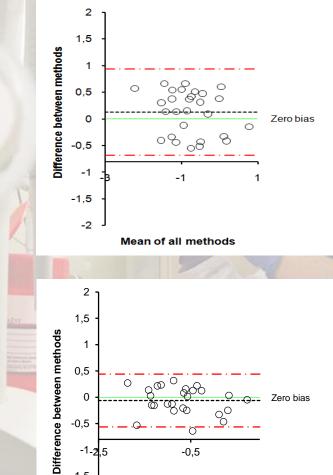


BCR-ABL monitoring in CML



BCR-ABL standarization





-0,5

Mean of all methods

-1-2.5

-1,5

-2

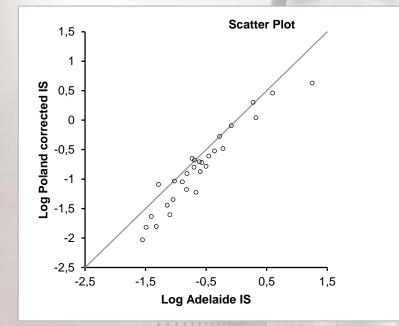
8/28 within 2-fold (29%) 19/28 within 3-fold (68%) 28/28 within 5-fold (100%)

21/26 within 2-fold (81%) 24/26 within 3-fold (92%) 26/26 within 5-fold (100%)



2011

BCR-ABL standarization



This one resulted in change of CF from 1 to 1,89

2016

EUTOS MR4.5

Data summary GUSB control gene : overall score and score per category

Lab Number	Final Score	10% cell line GUSB copy number	10% cell line %BCR-ABL IS value	CMR cell line GUSB copy number	CMR cell line BCR-ABL detected	conv	ABL detected	cDNA GUSB copy number	cDNA BCR-ABL copy number	cDNA ratio	Audit data
40											
12											
48											
37											
2											
41											
46											
25											
29											
38											
43											
1											

Scoring criteria:

Green = able to detect MR^{4.5} in a high proportion of samples

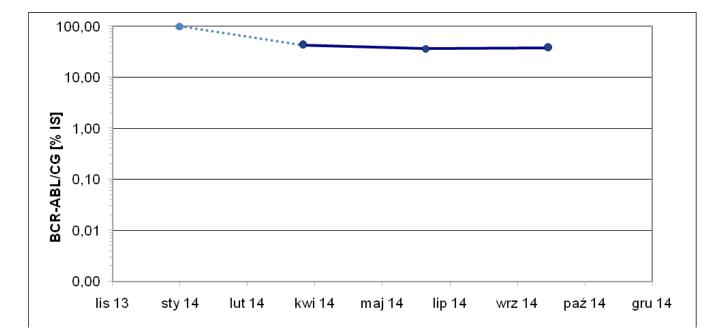
Orange = able to detect MR^{4.5} in a proportion of samples

Red = unable to detect MR^{4.5} in most samples



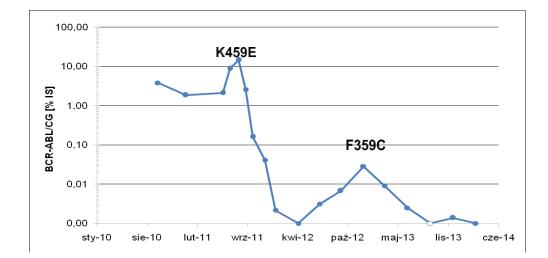
TKI resistance in CML

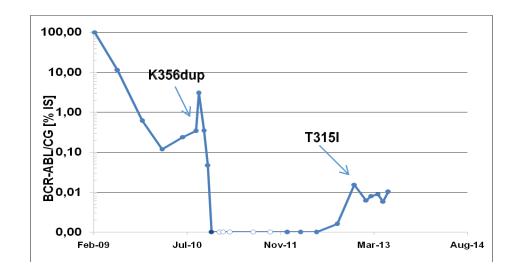
- Primary resistance no significant improvement after introduction of TKI therapy
 - Usually not related to BCR-ABL kinase domain mutations

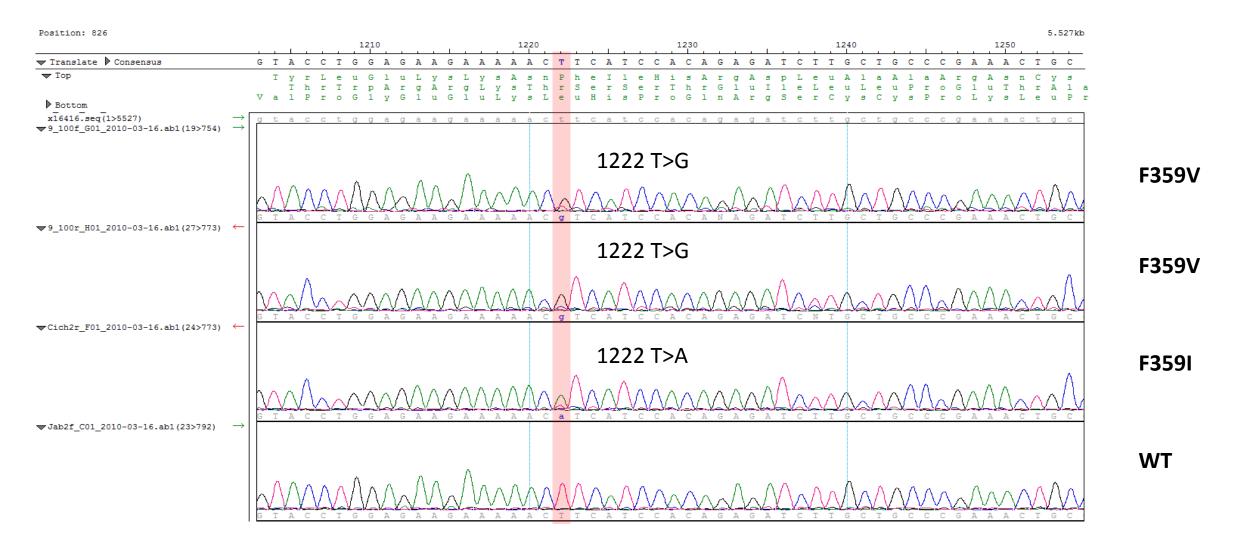


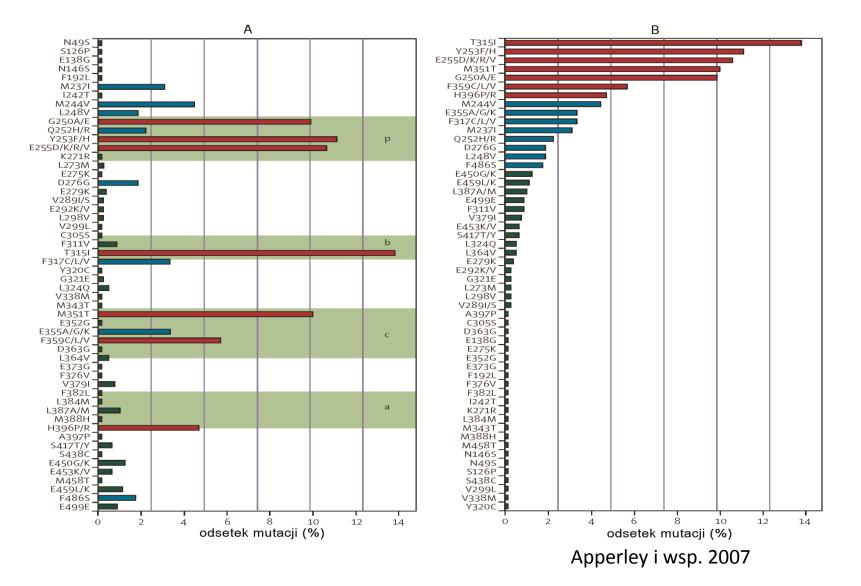
TKI resistance in CML

- Acquired resistance an increase of BCR-ABL transcript level after an initial reduction
 - Approx 50% of cases are related to BCR-ABL KD mutations
 - Before sequencing of BCR-ABL KD we have to be sure that patient has taken a full dose of TKI!









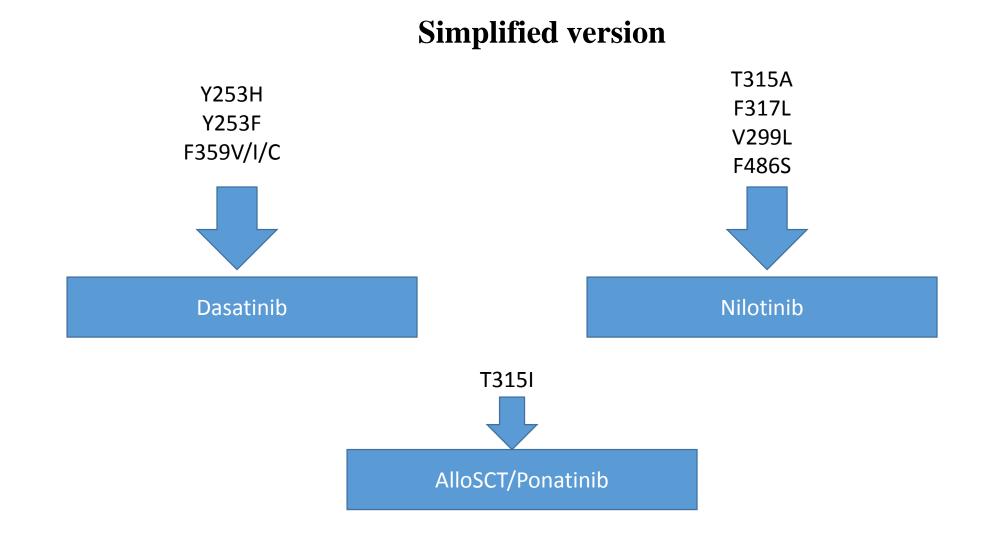
	Wartości Gl ₅₀ dla z			
	imatynib (nM)	nilotynib (nM)	dazatynib (nM)	
natywna forma BCR-ABL	260	13	0.8	
M244V	2000	38	1.3	
G250E	1350	48	1.8	
Q252H	1325	70	3.4	
Y253F	3475	125	1.4	
Y253H	>6400	450	1.3	
E255K	5200	200	5.6	
E255V	>6400	430	11	wrażliwy
V299L	540†	BD	18 [†]	średnio wrażli
F311L	480	23	1.3	
T315A	971	61	125†	niewrażliwy
T315I	>6400	>2000	>200	BD brak danych
F317L	1050	50	7.4	
F317V	350†	BD	53 [†]	
M351T	880	15	1.1	
E355G	2300 [‡]	BD	1.8 [§]	
F359V	1825	175	2.2	
V379I	1630	51	0.8	
L387M	1000	49	2	
H396P	850	41	0.6	
H396R	1750	41	1.3	

			zmiana IC ₅₀ (krotność, WT=1)									
		Zm	liana IC ₅₀ (Kr	otnosc, v	V I = 1)						
		bozutynib	imatynib	dazaty	nib	nilotyı	nib					
wт		1	1	1	1							
L248	V	2.97	3.54	5.11		2.80						
G250	Е	4.31	6.86	4.45	;	4.56	5					
Q252	Н	0.81	1.39	3.05	5	2.64	ł					
Y253	F	0.96	3.58	1.58		3.23	3					
E255	К	9.47	6.02	5.61		6.69)					
E255	V	5.53	16.99	3.44		10.31						
D276	G	0.60	2.18	1.44		2.00						
E279	К	0.95	3.55	1.64		2.05						
V299	L	26.10	1.54	8.65		1.34						
T315		45.42	17.50	75.03		39.4	1					
F317	L	2.42	2.60	4.46		2.22						
M351	Т	0.70	1.76	0.88		0.44						
F359	V	0.93	2.86	1.49		5.16						
L384	М	0.47	1.28	2.21		2.33						
H396	Ρ	0.43	2.43	1.07		2.41						
H396	R	0.81	3.91	1.63		3.10						
G398	R	1.16	0.35	0.69	0.69)					
F486	s	2.31	8.10	3.04		1.85	5					
	wra	żliwy										
	umi	arkowanie op	oorny		2	.01-4						
	оро	rny			4.0)1 - 10						

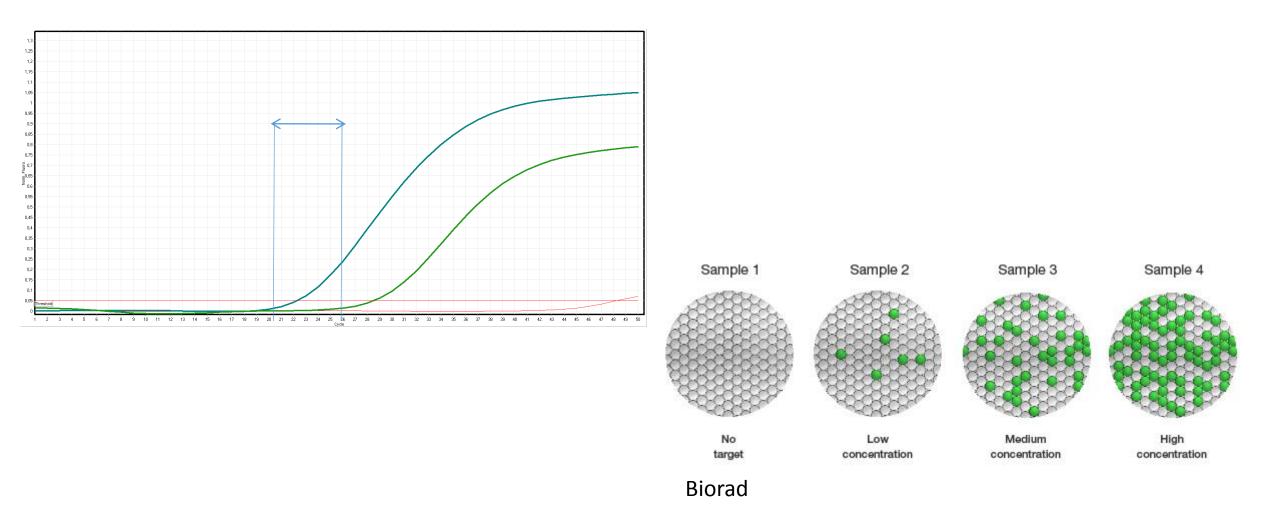
bardzo oporny

O'Hare i wsp. 2007, Redaelli i wsp. 2009

średnio wrażliwy



qPCR vs ddPCR



Sanger vs NGS

	214,400K 214,500K 214,600K 214,700K 214,800K 214,900K 215,000K 215,100K 215,100K <th< th=""></th<>
	1:237,200K 1:237,400K 1:237,600K 1:237,800K 1:238,
	RYR2 Translation 2950 2950 2960 K G E H F P Y E Q E I K F F A K G E H F P Y E Q E I K F F A H F P Y E Q/R E I K F F A
G G A G A A C T C C C T A T A G C G A T G G C T C T G G C	SNP db_xref ClinVarl2 Cosmiclv71
143185 143190 143195 143.2k 143205 143210	custom/ES
	Mutation Calls
GGAGAACTCCCTATAGSGATGGCTCTGGC	Reference 1:237,841,370 1:237,841,380 1:237,841,390 1:237,841,400 Pile-Up A A G G A A C A T T T C C C T T A T G A A C A A G A A A T C A A G T T C T T T G C A A A G G A G A A C A T T T C C C T T A T G A A C A G A A A T C A A G T T C T T T G C A A A G G A G A A C A T T T C C C T T A T G A A C G A G A A A T C A A G T T C T T T G C A A A G G A G A A C A T T T C C C T T A T G A A C G A G A A A T C A A G T T C T T T G C A A A G G A G A A C A T T T C C C T T A T G A A C G A G A A T C A A G T T C T T T G C A A A G G A G A A C A T T T C C C T T A T G A A C G A G A A A T C A A G T T C T T T G C A A A G G A G A A C A T T T C C C C T T A T G A A C G A G A A A T C A A G T T C T T T G C A A A G G A G A A C A T T T C C C C T T A T G A A C G A G A A A T C A A G T T C T T T G C A A A G G A G A A C A T T T C C C T T A T G A A C G A G A A A T C A A G T T C T T T G C A A A G G A G A A C A T T T C C C C T T A T G A A C G A G A A A T C A A G T T C T T T G C A A A G G A G A A C A T T T C C C C T T A T G A A C G A G A A A T C A A G T T C T T T G C A A A G G A G A A C A T T T C C C C T T A T G A A C G A G A A A T C A A G T T C T T T G C A A A G G A G A A C A T T T C C C C T T A T G A A C G A G A A A T C A A G T T C T T T G C A A A G G A G A A C A T T T C C C C T T A T G A A C G A G A A A T C A A G T T C T T T G C A A A G G A G A A C A T T T C C C C T T A T G A A C G A G A A A T C A A G T T C T T T G

NGS





Oxford Nanopore



ThermoFisher IonTorrent



Pacific Biosciences

Archer DX

ALL

ABL1		ABL2		AICDA		BCL11B		BCL2	BCL6		BCR	
BLNK		BRAF	•	CD274		CHD1		CREBBP	CRLF2		CSF1R	
CTLA4		DNM2	•	DNTT		EBF1		EPOR	ETV6		EZH2	•
FBXW7	•	FGFR1		FLT3		HOXA10		HOXA9	IDH1	•	IDH2	•
IKZF1		IKZF2		IKZF3		IL7R	•	IRF4	IRF8		JAK1	•
JAK2		JAK3	•	KDM6A	•	KLF2		KMT2A	KRAS	•	LMO1	
LYL1		MLLT4		MPL	•	MYC		NF1	NOTCH1		NRAS	•
NT5C2	•	NTRK3		NUP214		NUP98		P2RY8	PAG1		PAX5	
PBX1		PDCD1		PDCD1LG2		PDGFRA		PDGFRB	PICALM		PTK2B	
PTPN1		PTPN11	•	RAG1		RAG2		RUNX1	SEMA6A		SETD2	
SH2B3	•	SOX11		STAT3	•	STAT5B	•	STIL	TAL1		TCF3	
TLX1		TLX3		TYK2		WT1		ZCCHC7				

Legend:

SNV/Indel

Expression

Fusion, splicing or exon-skipping

* Internal tandem duplication (ITD)

CNV 🎆

Heme 2

ABL1	ABL2	ALK	BCL11B	BCL2	BCL3	BCL6	
BCR	BIRC3	CBFB	CCND1	CCND2	CCND3	CD274	
CDK6	CDKN2A	CEBPA	CEBPD	CEBPE	CEBPG	CHD1	
CHIC2	CIITA	CREBBP	CRLF2	CSF1R	CTLA4	DEK	
DUSP22	EBF1	EIF4A1	EPOR	ERG	ETV6	FGFR1	
FOXP1	GLIS2	ID4	IKZF1	IKZF2	IKZF3	IRF4	
IRF8	JAK2	KAT6A	KLF2	KMT2A	MALT1	MECOM	
MKL1	MLF1	MLLT10	MLLT4	MUC1	MYC	MYH11	
NF1	NFKB2	NOTCH1	NTRK3	NUP214	NUP98	P2RY8	
PAG1	PAX5	PDCD1	PDCD1LG2	PDGFRA	PDGFRB	PICALM	
PML	PRDM16	PTK2B	RARA	RBM15	ROS1	RUNX1	
RUNX1T1	SEMA6A	SETD2	STIL	TAL1	TCF3	TFG	
TP63	TYK2	ZCCHC7					

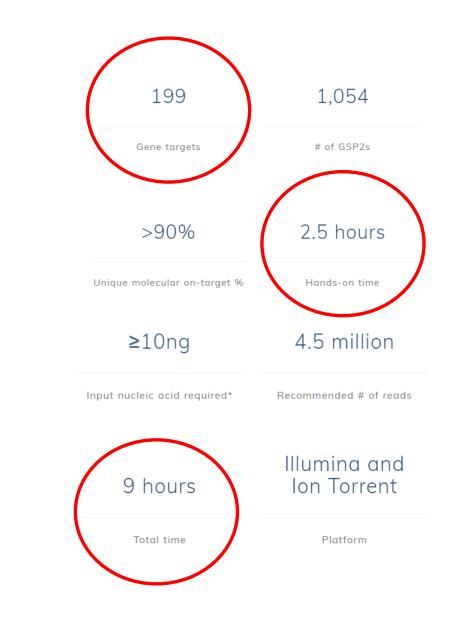
Legend:

SNV/Indel
 Expression
 Fusion, splicing or exon-skipping
 Internal tandem duplication (ITD)

CNV

Archer DX – PAN heme

ABL1		ABL2		AICDA		АКТЗ	•	ALK		ASB13		ASXL1	•	
BATF3		BAX	•	BCL11B		BCL2		BCL2A1		BCL3		BCL6		
BCR		BIRC3		BLNK		BMF		BMP7		BRAF	•	ВТК	•	
CALR	•	CARD11	•	CBFB		CBL	•	CCDC50		CCND1		CCND2		
CCND3		CD274		CD44		CD79B	•	CDC25A		CDK6		CDKN2A		
CDKN2B		CEBPA		CEBPD		CEBPE		CEBPG		CHD1		CHIC2		
CIITA		CREB3L2		CREBBP		CRLF2		CSF1R		CSF3R	•	CTLA4		
CYB5R2		DCK	•	DEK		DENND3		DLEU1		DNM2	•	DNMT3A	•	
DNMT3B		DNTT		DUSP22		E2F2		EBF1		EIF4A1		ENTPD1		
EPOR		ERG		ETV6		EXOC2		EZH2	•	FAM216A		FBXW7	•	
FGFR1		FGFR2	•	FGFR3	•	FLT3		FOXP1		FUT8		GATA1		
GATA2	•	GLIS2		GNAS		HOXA10		HOXA9		ID4		IDH1		
IDH2	•	IKZF1		IKZF2		IKZF3		IL16		IL7R	•	IRF4		
IRF8		ІТРКВ		JAK1		JAK2		ЈАКЗ	•	KAT6A		KDM6A	•	
KIAA0101		KIT	•	KLF2		KMT2A		KRAS		LIMD1		LMO1		
LMO2		LRMP		LYL1		LZTS1		MAL		MALT1		MAML3		
MECOM		MKL1		MLF1		MLLT10		MLLT4		MME		MPL	•	
MUC1		MYBL1		MYC		MYD88	•	MYH11		NEK6		NF1		
NFKB1		NFKB2		NME1		NOTCH1		NOTCH2	•	NPM1	•	NRAS	•	
NT5C2	•	NTRK3		NUP214		NUP98		P2RY8		PAG1		PAICS		
PAX5		PBX1		PDCD1		PDCD1LG2		PDGFRA		PDGFRB		PHF6	•	
PICALM		PIM1		PIM2		PLCG1	•	PLCG2	•	PML		PPAT		
PRDM16		PRKAR2B		PTK2B		PTPN1		PTPN11	•	PYCR1		RAB29		Legend:
RAG1		RAG2		RANBP1		RARA		RBM15		RHOA	•	ROS1		
RUNX1		RUNX1T1		S1PR2		SEMA6A		SERPINA9		SETBP1	•	SETD2		 SNV/Indel Expression
SF3B1	•	SH2B3	•	SH3BP5		SLC29A1	•	SOX11		SRSF2	•	STAT3	•	 Expression Fusion, splicing or exon-skipping
STAT5B	•	STAT6	•	STIL		STRBP		TAL1		TCF3		TFG		Internal tandem duplication (ITD) CNV
TLX1		TLX3		TNFRSF13B		TNFSF4		TP63		TYK2		U2AF1	•	III CINV
WT1		XPO1	•	ZCCHC7										



VDJ rearangements

•Mechanism of recombination which occurs on early stage of lymphocyte development

•Plays role in forming of antibodies (B-cells) and T-cell receptors (TCR)

•VDJ recombination – name derived from regions participating in rearrangement

•V – variable, D – diversity, J – joining

is required for this analyses

In lymphoid malignancies, it can be used as the clonality marker
Clonal rearrangements can be detected with molecular techniques
Once detected, certain rearrangement pattern can be used for
MRD monitoring with ultimate sensitivity
Due to the complexity of the problem, highly specialized laboratory

V-gene Region D-gene Region J-gene Region D-J Rearrangement V-DJ Rearrangement Transcription/Translation